Data Sheet

Streptomycin (SM) ELISA Kit

Cat. #SG-4011

Size: 96 Wells

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Principle and Application

This kit adopts the method of indirect competitive enzyme-linked immunoassay (ELISA) to detect Streptomycin (SM) in tissue, honey and other samples. The kit is composed of Microtiter Plate coated with coupled antigens, HRP conjugate, antibodies, standards and other supporting reagents. During the detection, with adding standards or samples, the SM in the samples will compete with the coupled antigens to combine with anti-SM antibodies. After adding HRP conjugate, take coloration with TMB substrates. Absorbance value of the samples is a negative correlation with SM content. Lastly, by comparing the obtained absorbance values with the standard curve, we can calculate the SM content in the sample.

Storage conditions

- The kit shall be stored at 2-8 ℃. Avoid freezing.
- Shelf life: 12 months. The date of manufacture is presented in the label of the box.

Technique Data

- Kit Sensitivity: 0.1ppb (ng/mL)
- Reactive Mode: 25℃, 30min～30min～15min
- Detection Limits:

Sample	Detection Limits
Tissue	4ppb
Honey, royal jelly	2ppb
Milk, milk powder	5ppb
Poultry eggs	10ppb

- Cross-reaction Rate:

Streptomycin……100%

Dihydrostreptomycin ……...100%

Kanamycin……6.3%

Gentamicin……2.5%

- Sample Recovery Rate:

Sample	Recovery Rate
Tissue, honey, royal jelly, milk, milk powder， eggs	85±15%

Composition of the Kit

Reagent	Specification
Microtiter Plate	8wells× 12strips
Standard: 0ppb, 0.1ppb, 0.3ppb, 0.9ppb, 2.7ppb, 8.1ppb	1.0mL each
High Standard (red cap)：1ppm	1×1.0mL
Antibody solution (blue cap)	1×5.5mL
HRP conjugate (red cap)	1×11mL
Substrate Reagent A (white cap)	1×6mL
Substrate Reagent B (black cap)	1×6mL
Stop Solution (yellow cap)	1×6mL
Concentrated Wash Buffer (20×)(white cap)	1×40mL
Concentrated Reconstitution Buffer (5×)(yellow cap)	1×50mL
Instructions	1
Adhesive Membrane	1
Sealed bag	1

Materials Required but Not Supplied

- **Equipment:** microplate reader, printer, grinder (for homogenizing solid samples), nitrogen evaporator, vortex mixer (**for shake and mix**), centrifuge, graduated transfer pipette, and balance with a division value of 0.01 g, constant temperature device(25°C), water bath;

- **Micropipette:** single-channel (20-200μL and 100-1000μL), and multi-channel 300μL;
- **Reagents:** NaOH, $Na_2HPO_4 \cdot 12H_2O$, $NaH_2PO_4 \cdot 2H_2O$, N-hexane, concentrated phosphoric acid (85%w/w), Acetic acid, Methanol.

Experimental preparation

Restore all reagents and samples to room temperature (adjust to around 25°C) for more than 30 min before use. This is a crucial step to ensure there is no precipitation in the reagents.

Please note that the labware must be clean. Use disposable pipette tips to avoid contamination of interference results.

◆ **Solution preparation:**

Solution 1: 0.05M Phosphate Buffer

Weigh 12.9g of $Na_2HPO_4 \cdot 12H_2O$ and 2.175g of $NaH_2PO_4 \cdot 2H_2O$. Dissolve in deionized water and mix thoroughly. Bring the volume up to 1000mL.

Solution 2: 0.04M Phosphoric Acid Solution (for honey, royal jelly)

Add 1 mL of concentrated phosphoric acid(85%w/w) to deionized water, mix thoroughly, and bring the volume up to 360 mL.

Solution 3: 1M NaOH Solution (for honey, royal jelly)

Weigh 4g of NaOH, dissolve in deionized water, mix thoroughly, and bring the volume up to 100mL.

Solution 4: 1% Acetic Acid Solution (for poultry egg)

Measure 1 mL of acetic acid and add to 99 mL of deionized water, mix thoroughly to dissolve.

Solution 5: 70% Methanol Solution (for Poultry egg)

Measure 700 mL of methanol and add to 300 mL of deionized water, mix thoroughly to dissolve.

Solution 6: Reconstitution Buffer

Dilute the Concentrated Reconstitution Buffer (5×) five times (Concentrated Reconstitution Buffer (5×) /Deionized water= 1:4) .The Reconstitution Buffer can be stored for one month at 4°C.

Solution 7: Working Wash Buffer

Dilute the concentrated wash buffer (20**×)** by a factor of 20,

(Concentrated wash buffer/Deionized water= 1: 19)

◆ **Sample pretreatment steps:**

1. Tissue treatment.

1) Weigh 2±0.05g of defatted homogenized sample. Add 8 mL of **0.05M Phosphate Buffer (Solution 1)**. Vortex for 5 minutes and then incubate in a 56°C water bath for 30 minutes.

2) Centrifuge at room temperature at 4000 rpm or higher for 10 minutes.

3) Take 1 mL of the supernatant and add 1 mL of **N-hexane**. Mix thoroughly. Centrifuge at room temperature at 4000 rpm or higher for 5 minutes.

4) Remove the upper layer. Take 50μL of the lower layer and add 450μL of the **Reconstitution Buffer (Solution 6)**. Mix for 30 seconds.

5) Take out 50μL for test.

Dilution times of the sample:40 ***Detection limits: 4ppb***

2. Honey and Royal Jelly treatment.

1) Weigh 2±0.05g of the sample and add 4 mL of **0.04M Phosphoric Acid Solution (Solution 2)**. Vortex until completely dissolved. Centrifuge at room temperature at 4000 rpm or higher for 5 minutes until clear. *(For honey samples, centrifugation can be skipped and proceed directly to Step 2.)*

2) Add 450μL of **1M NaOH Solution (Solution 3)** to adjust the pH to between 7 and 9. (For royal jelly, transfer all of the supernatant to a clean centrifuge tube and adjust the pH to

between 7 and 9.) Centrifuge at room temperature at 4000 rpm or higher for 5 minutes until clear.

3) Take 50μL of the supernatant and add 450μL of the **Reconstitution Buffer (Solution 6)**. Mix thoroughly for 30 seconds.

4) Take out 50μL for test.

Dilution times of the sample:20 Detection limits: 2ppb

3. Milk and milk powder treatment.

1) Weigh 2±0.05g of the sample and add 8 mL of **0.05M Phosphate Buffer (Solution 1)**. Vortex for 5 minutes and then incubate in a 56°C water bath for 30 minutes.

2) Centrifuge at room temperature at 4000 rpm or higher for 10 minutes.

3) Remove the upper fat layer. Take 50μL of the middle clear layer and add 450μL of the **Reconstitution Buffer (Solution 6)**. Mix thoroughly for 30 seconds.

4) Take out 50μL for test.

Dilution times of the sample:50 Detection limits: 5ppb

4. Poultry egg treatment.

1) Peel and homogenize the poultry egg. Weigh 1g ± 0.05g into a 50 mL centrifuge tube. First, add 2 mL of **1% Acetic Acid Solution (Solution 4)** and vortex for 2 minutes. Then add 7 mL of **70% Methanol Solution (Solution 5)** and vortex for another 2 minutes. Centrifuge at room temperature at 4000 rpm for 10 minutes.

2) Transfer 0.1 mL of the upper layer into a 1.5 mL centrifuge tube. Add 0.9 mL of **Reconstitution Buffer (Solution 6)** and vortex for 30 seconds.

3) Take out 50μL for test.

Dilution times of the sample:100 Detection limits: 10ppb

ELISA procedure

Place all reagents and samples to room temperature (adjust to around 25°C) for 30min.

Gently shake the reagent bottles before use.

Take out the frame of the microplate along with the required number of wells. Then place the unused microplate wells into the sealed bag with the desiccant provided. Store the remaining kit in the refrigerator at 2-8°C.

Step 1: Number: Number the wells in sequence corresponding to the samples and standard, make 2-well parallel trials for each sample and standard, and record their locations.

Step 2: Sample Incubation: Add 50μL of **standard or sample** into each numbered well, then add 50μL of **antibody solution** into each well. Finally, cover the Microtiter Plate with the adhesive membrane, shake gently by hand (or use a microplate shaker) for 5s and incubate for 30 min at 25℃ in the dark.

Step 3: Washing: Uncover the adhesive membrane carefully, remove the liquid, pipette 350μL of **Working Wash Buffer (Solution 7)** to every well, let stand for 30 seconds then drain, repeat 5 times. Invert the plate and tap it against a thick absorbent paper (or lint-free cloth), with a soft towel placed underneath. (Bubbles that are not removed after tapping dry can be punctured with a clean pipette tip).

Step 4: Enzyme Incubation: Add 100μL of **HRP conjugate** into each well. Then cover the Microtiter Plate with the adhesive membrane, incubate for 30 min at 25℃ in the dark.

Step 5: Washing: Same as step 3.

Step 6: Color: Add 50μL of **Substrate Reagent A** to each well. Then add 50μL of **Substrate Reagent B** per well. Shake gently by hand (or use a microplate shaker) for 5s, and allow to react for 15min at 25℃ in the dark. (The reaction can be extended appropriately if the blue color is too pale.)

Step 7: Stop the reaction: Pipette 50μL of **Stop Solution** to each well, and shake gently by hand (or use a microplate shaker). The reaction would be stopped.

Step 8: Calculate: Determine the Optical Density (OD value; absorbance value) at 450nm (Reference wavelength 630nm) with a microplate reader. Finish this step within 10min after stop the reaction.

Interpretation of result

◆ **Calculate the percentage of absorbance value**

$$\textbf{Percentage of absorbance value}(\%) = \frac{A}{A0} \times 100\%$$

A—the average OD value of the sample or the standard;

A0—the average OD value of the 0ppb standard.

It is used to calculate the percentage absorbance of a standard or sample.

◆ **Draw the standard curve and calculate**

- Take absorbance percentage(A/A0) of standards as Y-axis and the corresponding log of standards concentration (ppb) as X-axis.
- Draw the standard semi-log curves with X-axis and Y-axis.
- Take absorbance percentage of samples substitute into standard curve, then can get the corresponding concentration from standard curve. **Last, the resulting concentration values multiplied by the corresponding dilution times is the actual concentration of SM of samples.**

  If professional analysis software of the kit is used for calculation, it is more convenient for accurate and rapid analysis of a large number of samples.

Attention

- Before test, the reagents and samples should be balanced to room temperature (25℃). If below 25℃, it will lead to all the standard OD value on the low side.
- In washing process, dry wells may result in non-linear standard curves and undesirable reproducibility. Therefore, proceed to the next step immediately after washing.

- Please mix the contents within the wells uniformly and wash the plate thoroughly. The reproducibility is largely determined by consistency of washing step.
- During the incubation, cover microplates with adhesive membrane to avoid light.
- Do not use kits that are overdue. Do not mix reagents with those from other lots.
- Substrate Reagent A/B is colorless. If not, please discard.
- If absorbance value of 0ppb is below 0.5（A450nm< 0.5 ）, it means that the reagent may be metamorphic.
- Stop solution is corrosives. Please avoid contacting with skin.
- **As the OD values of the standard curve may vary according to the conditions of actual assay performance (e.g. operator, pipetting technique, washing technique or temperature effects), the operator should establish a standard curve for each test.**
- **For the mentioned sample, fast and efficient extraction methods are included in the kit description. Please consult technical support for the applicability if other sample need to be tested.**
- The kit is used for rapid screening of actual samples. If the test result is positive, the instrument method such as HPLC, LC/MS can be used for quantitative confirmation.